

# Extended Protocol

## RapidPro Spin™

### Protein Purification Kit

QCD-025

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\*All changes must be recorded. This is a master, living, document. Printed copies/PDFs are not controlled.

### Kit Components

1. 125 mL Proteios **L1** Lysis Buffer (Qty 1)
2. 150 mL Proteios **W2** Wash Buffer (Qty 1)
3. 125 mL Proteios **E1** Elution Buffer (Qty 1)
4. Pre-packed dry Spin Columns (Qty 8)

*\*Sterile 50 mL Tubes and Centrifuge required.*

## Introduction

Proteios RapidPro Spin™ purification kit is based on the interaction between the Proteios Car9 affinity tag and the Proteios RapidPro Spin™ Resin.

The Proteios RapidPro Spin™ Purification Kit has been optimized to provide high-purity isolation of very low- to moderate-expressed proteins in *E. coli*.

**Purification takes about 20 minutes with an average purity > 90%.**

The target protein of interest (POI) can be expressed with a Proteios Car9 tag at either the N-terminal or C-terminal position. Proteios Car9-tagged POI binds to the Proteios RapidPro Spin™ Resin and can be released with the addition of the Proteios Elution Buffer.



# Cloning

The target protein of interest (POI) can be expressed with a Proteios Car9 tag at either the N-terminal or C-terminal position. Proteios Car9-tagged POI binds to the Proteios RapidPro Spin™ Resin and can be released with the addition of the Proteios Elution Buffer.

**Proteios Car9 DNA sequence** (GATAGCGCACGCGGTTTCAAAAACCGGGTAAACGC) coding for DSARGFKKPGKR and the Proteios' spacer (AAGCTTGGCGGCGGCTCT) coding for GGS can be introduced at the N-terminal of the protein of interest after the start codon ATG.



Alternatively, they can be introduced to the C-terminal before any stop codon(s) – first the spacer (AAGCTTGGCGGCGGCTCT) and then the Proteios Car9 coding sequence (GATAGCGCACGCGGTTTCAAAAACCGGGTAAACGC).



In addition to the spacer reported above, other spacers might be introduced between the CDS (Coding Sequence) of the protein of interest and the Proteios Car9 CDS.

We recommend gene synthesis and subcloning to insert both Proteios Car9 and Spacer CDS in frame with the protein of interest CDS. Alternatively, a PCR-based approach can be used to insert both Proteios Car9 and spacer CDS in the desired position prior to the subcloning of the new sequence in the desired expression plasmid.

We strongly recommend to sequence verify the insert sequence after cloning into your preferred expression plasmid.

The sequence verified expression plasmid may be transformed into Top10 chemically competent cells (Thermo Fisher) or analog cell line to amplify plasmid DNA according to manufacturing instructions. Commercially available plasmid purification kits can be used to extract and purify the plasmid DNA.



# Cell Culture

BL21(DE3) or similar cell lines can be used to express the POI according to manufacturing instructions. Proteios has successfully tested them with both IPTG and arabinose induction systems. Usually, 1 mM IPTG or 0.1% arabinose are enough to induce protein expression.

1. Transform BL21(DE3) or similar competent cell lines according to the manufacturer's instruction.
2. Plate the transformed bacteria on agar Petri plates with appropriate antibiotics to obtain about 100 colonies on the plate.
3. Grow colonies overnight at 37°C.
4. The next day, pick a colony and grow it overnight in 5 mL of LB media at 37°C in a shaking incubator at 200 rpm. Addition of appropriate antibiotics is recommended.
5. The next morning, add 1 mL of overnight culture to 50 mL of media and incubate at 37°C in a shaking incubator at 200 rpm. Addition of appropriate antibiotics is recommended.
6. Grow bacteria until they reach the desired OD600.
7. Add IPTG or Arabinose to induce protein expression (other induction systems can be used).
8. Grow cells for the required time and temperature to allow the expression of the POI.
9. Harvest the cells by centrifugation, remove supernatant and either proceed to the next step or freeze cell pellets and store at -20°C.



# Cell Lysis

Proteios Lysis Buffer does not contain: EDTA, DNase, reducing agents and protease inhibitors.

*You can supplement the Proteios' Buffers with the desired amounts of these supplements as needed.*

Proteios Lysis Buffer has been tested with high-pressure homogenization, sonication and freeze/thaw.

Resuspend the cells in Proteios Lysis buffer;

4 mL of Lysis buffer is recommended for a pellet obtained from a 50 mL bacteria culture (about 10 mL of lysis buffer /g of bacteria pellet).

To lyse bacteria, **use 1** of these 4 recommended methods:

1. Microfluidizer homogenizer
2. High-pressure homogenizer: 3 passages on French Press, 540 psi
3. Sonication: 6 cycles of sonication on ice bath by alternating 5 seconds on and 30 seconds off at 30% power
4. Freeze/thaw: 3 cycles of freeze and thaw by alternating -80°C and 37°C.  
Note: Freeze at -80°C until solid for about 5 minutes, then thaw at 37°C until liquid, vortex before next freezing step.

Depending on the target protein, one method may be more suitable than the others.

*(Usually freeze/thaw leads to a lower yield and higher purity.)*

Centrifuge the cell lysate at 10000 x g for 15 minutes at 4°C.

**The supernatant can be directly loaded on the equilibrated RapidPro Spin™ column.**



# Quick Protocol **RapidPro Spin™**

QCD-027 **Intended use**-For research use only.

Revision Date: 2023 July 27

## Before Starting

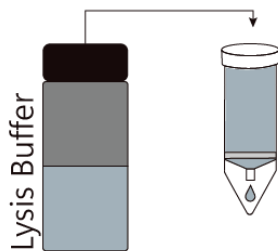
- Cloning
- Cell Culture
- Cell lysis. Thaw lysate.

See [Extended Protocol for Details](#).

## Kit Components

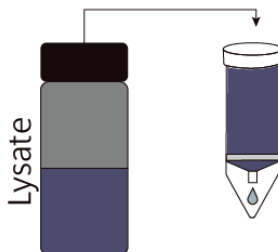
- 125 mL Proteios L1 Lysis Buffer (Qty 1)
- 150 mL Proteios W2 Wash Buffer (Qty 1)
- 125 mL Proteios E1 Elution Buffer (Qty 1)
- Pre-packed dry Spin Columns (Qty 8)

## Purification steps



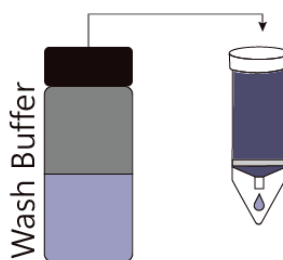
### 1. Equilibration: Equilibrate the column.

- Proteios RapidPro Spin™ Columns are shipped dry and should be equilibrated prior to adding the clarified lysate.
- Insert dry column into empty sterile 50 mL tube.
- Add 4 mL of Proteios RapidPro Spin™ Lysis Buffer to the column.
- Cap, then centrifuge in 50 mL tube at 500 x g for 3 minutes at room temperature. Discard the flow through (FT).



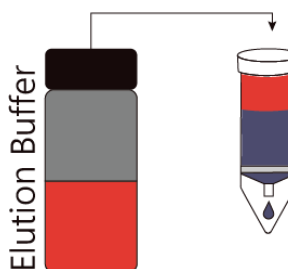
### 2. Binding: Load Lysate on the column.

- (**Recommended**: save from 20 µL to 40 µL of lysate for SDS gel.)
- Load 4 mL of lysate to the column, cap, and centrifuge the column at 500 x g for 3 minutes at room temperature.
- Discard the FT.



### 3. Washing: Wash the column.

- Wash the column with 4 mL of Proteios RapidPro Spin™ Wash Buffer two times.
- Centrifuge at 500 x g for 3 minutes at room temperature for each wash.
- Discard the FT.



### 4. Elution: Elute the Protein of Interest.

- Replace the 50 mL tube for elution step.
- Add 4 mL Proteios RapidPro Spin™ Elution Buffer to the column.
- Centrifuge at 500 x g for 3 minutes at room temperature.
- **The Protein of interest (POI) is in the eluate.**
- Repeat Elution step one more time to recover the remaining Proteios Car9-tagged POI.



# Modifications/Troubleshooting

Problem	Suggestion
POI does not bind the matrix	<ul style="list-style-type: none"> <li>• Add up to 500 mM NaCl to the Lysis Buffer.</li> <li>• Add up to 10% glycerol and/or up to 0.3% Tween 20 to the Lysis Buffer.</li> <li>• Extend the length of the spacer between Proteios Car9 and POI.</li> </ul>
POI binds the matrix, but it is eluted during washes	<ul style="list-style-type: none"> <li>• Lysis Buffer can be used to perform the washes instead of Wash Buffer.</li> <li>• A mixture of Lysis Buffer and Wash Buffer can be used to find the optimal purity/yield balance.</li> </ul>
POI purity is < 80%	<ul style="list-style-type: none"> <li>• Lyse cells using freeze/thaw.</li> </ul>

